

## # Evaluations

①

- ① Sterility test
- ② Clarity test
- ③ leakage test
- ④ pyrogen test
- ⑤ AMay.

### ① Sterility test :-

It is done by detecting the presence of viable forms of bacteria, fungi and yeast in parenteral products.

The test for sterility must be carried out under aseptic conditions in order to avoid accidental contamination of the product during test.

②

∴ All glassware required for the test must be sterile.

→ The test for sterility may be carried out either by

- (a) Membrane filtration method
- (b) Direct inoculation method

Principle - If bacteria or fungi are placed in a medium which provide nutrition + water, + kept at a favourable temperature, organism will grow + their presence can be indicated by their turbidity in the clear ~~st~~ solution.

→ Steps involved in sterility testing

- I) Selection of the sample
- II) Selection of the quantity of product to be used
- III) method of testing
- IV) observation + results.

→ Selection of the Sample

Sample must be representative of the whole of bulk material + lot of final containers.

∴ Random sampling is taken from final containers

| No. of items in the batch      | Minimum no. of items recommended to be tested |
|--------------------------------|---|
| → not more than 100 containers | 10% or 4 containers whichever is greater      |
| → between 100 to 500 "         | 10 Containers                                 |
| → more than 500 containers.    | 2% or 20 Container whichever is less          |

⇒ Selection of the quantity of products to be used

↳ Depends mainly on the volume or weight of the containers.

↳ Minimum samples to be used in each culture medium in the test for sterility are

| Quantity of each container             | Minimum quantity to be used in each culture medium |
|--|--|
| $L < 1 \text{ ml}$                     | whole contents of container                        |
| $L = 1 \text{ ml or } < 40 \text{ ml}$ | $\frac{1}{2}$ content of "                         |
| $L \geq 100 \text{ ml or more}$        | 10% content of "                                   |

⇒ methods of testing

(a) membrane filtration method

(b) direct inoculation method.

Membrane filtration method (method-1)

∴ This method is suitable for

→ Filterable aqueous preparations.

→ Alcoholic preparations

→ oily preparations.

②

Media used for membrane filtration method

(A) Fluid Thioglycolate Media

primarily intended for the culture of aerobic + anaerobic bacteria

→ Incubation of media: 14 days at 30-35°C

(B) Soyabean Casein digest medium primarily intended for the culture of fungi.

∴ Incubation of media: 14 days at 20-25°C

∴ Steps

membrane filter 0.45 μ porosity



Filter the test solution



After filtration remove the filter



Cut the filter in two halves.

First halves (for Bacteria)



Transfer in 100 ml culture media / Fluid thioglycolate medium



Incubate at 30-35°C for not less than 7 days.



Observe the growth in the media

Second halves (for fungi)



Transfer in 100 ml culture media (Soyabean-casein digest media)



Incubate at 20-25°C for not less than 7 days



Observe the growth in media.

→ Direct inoculation method (method-2) :-

:- Suitable for samples with small volumes.

:- Volume of the product is not more than 10% of the volume of media

:- Suitable method for aqueous solutions, oily liquids, ointments and creams.

:- Direct inoculation of the culture medium suitable quantity of the preparation to be examined is transferred directly into the appropriate culture medium & incubate for not less than 14 days.

### Observation & Results

:- culture media is examined during and after at the end of incubation. The following observations are possible.

→ No evidence of growth → Passed the test for sterility

→ There is evidence of growth → Re-testing is performed for same no. of sample, volume of media as in original test → no evidence of growth → Pass the test

→ There is evidence of growth → Isolate & identify the organism.

→ Re-testing is performed with twice no. of sample.

## ② Clarity test

It is performed to ensure that parenterals are free from visible foreign particles.

Each parenteral preparation in its final container is subjected individually to a visual inspection to exclude the possibility of foreign particles.

The contents of the container are held by the neck against strongly tilted slowly inverted & rotated, then examined. It may be dangerous when the particle size is larger than R.B.C. & may block the blood vessel. This type of products are immediately rejected from the batch.

Applicable for :- 100 ml or more Volume Contained of single dose IV given by IV infusion

Not applicable for :- multi-dose injections Single dose SVP Injectable Solutions constituted from Sterile Solids.

Methods of monitoring particulate matter Contamination :-

- ① Visual method
- ② Coulter Counter method
- ③ Filtration method
- ④ Light blockage method.

②

### ③ Seal Packaging / Leaking test

!- The sealed ampoules are subjected to small cracks which occur due to rapid temperature changes or due to mechanical shocks.

Filled & sealed ampoules.



Dipped in 1% methylene blue solution under negative pressure in vacuum chamber



Vacuum released coloured solution enter into the ampoule.



Defective Sealing. [vials & bottles are not suitable for this test because the sealing material used is not rigid]

### ④ Pyrogen testing

!- Pyrogen = Pyro (fire) + gen (beginning)

!- Fever producing, metabolic by-products of microbial growth and death.

#### ⇒ (A) Rabbit test

Rabbits are used to perform this test because their body temp. increases when pyrogen are introduced

in to their bodies by Parenteral route.

!- 3 healthy adult rabbits of either sex each weighing NLT 1.5 Kg are selected

!- Do not use any rabbit +  
→ having a temp > higher than 39.8°C  
→ Showing temp. Variation > 0.2°C b/w two successive reading

!→ method

Dissolve the substance being examined in, or dilute it with a pyrogen free Saline Solution.

!- warm the liquid being examined to approx 38.5°C temp before injection.

!- The volume of injection is NLT 0.5 ml/kg + NMT 10 ml / Kg of body weight.

!- with held water during test

!- Clinical thermometer is inserted into the rectum of rabbit to record body temp.

!- 2 normal reading of rectal temp are should be taken prior to the test injection at an interval of 1/2 hr (30 min) + it's mean is calculated initial temp.



①  
:- The Sol<sup>n</sup> under test is injected through an ear vein.

:- Record the temp. of each rabbit in an interval of 30 min for 3 hrs.

:- The difference b/w initial temp & maximum temp is recorded -

Result

| <u>Rabbits.</u>                                   | <u>Individual Temp. (°C)</u> | <u>Temp. group (°C)</u> | <u>Test</u> |
|---|------------------------------|-------------------------|-------------|
| :- 3 Rabbits                                      | 0.6                          | 1.4                     | Passed      |
| :- 9/10 above test not passed<br>3+5 Rabbit taken | 0.6                          | 3.7                     | Passed      |

(B) LAL (Limulus Amoebocyte Lysate) test.

⑤ Assay

Assay is performed according to method in monograph.

:- Assay is done to check the quantity of medicament present in the preparation